

Effects of dried horse radish (*Moringa oleifera*) pod meal as feed additive on egg laying and performance of shaver brown birds

Ikpe JN¹, Ahaotu EO², Akomaegbe C³, Simeon-ahaotu VC⁴, Oko EC¹ and Ikwuagwu VO⁴

¹Department of Animal Production Technology, Akanu Ibiam Federal Polytechnic Unwana Afikpo, Ebonyi State, Nigeria

²Department of Animal Science, University of Agriculture and Environmental Sciences Umuagwo, Imo State, Nigeria

³Department of Animal Production and Health Technology, Imo State Polytechnic Omuma Campus, Nigeria

⁴Department of Microbiology, Gregory University, Uturu, Abia State, Nigeria

Corresponding author: juliananikpe@gmail.com

Received on: 10/08/2023

Accepted on: 21/11/2023

Published on: 27/11/2023

ABSTRACT

Aim: Main purpose of the study was to explore the effect of *Moringa oleifera* pods meal (MOPM) as feed additive on performance, carcass and egg laying of shaver brown birds.

Method and materials: Sixty shaver brown laying birds having average weight of 1.58±3.02 Kg were assigned to four treatments with three replicates and five birds per replicate in a completely randomized design. Layer diets were added with four levels (0, 5.00, 10.0 and 20.0% of MOPM).

Results: Results revealed that performance was improved as feed conversion ratio (FCR) and feed intake (FI) were decreased with the increase in inclusion levels of MOPM ($P \leq 0.05$).

Conclusion: It was concluded that *Moringa oleifera* pod meal may positively improve performance and egg laying percentage of layers.

Keywords: *Moringa oleifera* Pod Meal, Shaver Brown Laying Birds, Performance, Carcass, Additive.

Cite This Article as: Ikpe JN, Ahaotu EO, Akomaegbe C, Simeon-ahaotu VC, Oko EC and Ikwuagwu VO (2023). Effects of dried horse radish (*Moringa oleifera*) pod meal as feed additive on egg laying and performance of shaver brown birds. J. Vet. Res. Adv., 05(02): 68-77.

Introduction

A well formulated diet adequate in all nutrients including protein is essential for normal growth and physiological function of poultry (Onu, 2011). This is done by carefully balancing the nutrients involved and taking into account the physiological state of the bird (Abbas, 2013).

Soyabean meal and fish meal have been widely and successfully used as conventional protein sources for livestock production (Aniebo, 2013). However, the prices of these protein sources have been escalating continuously in recent times, whilst availability is often erratic. The problem has been worsened by the increasing competition between humans (demands for soybean) and livestock for these protein ingredients (Aniebo, 2013). Conventional synthetic feed additives such as antibiotic growth promoters, antioxidants, anti-parasitic agents and anti-fungal agents have been used in poultry feed for decades (Uduji *et al.*, 2020).

However, they created multiple complications, such as traceability in animal products and resistance to antibiotics in the consumer, which became public health issues (Embuscado, 2015). On these grounds, the use of all kinds of antibiotic growth promoters was banned in animal feed in Europe (Nkukwana *et al.*, 2014). Revolutions in animal feed production gave rise to the idea of phytogenic feed additives (Grashorn, 2010 and Odoemelam *et al.*, 2020). The recommended policy is to identify and use locally available feed resources to formulate diets that are as balanced as possible (Kakengi *et al.*, 2007). There is the need, therefore, to explore the use of non-conventional feed sources that have the capacity to yield the same output as conventional feeds and perhaps at cheaper cost. Hence, any similar high protein ingredient which could partially or completely be used as a substitute for soyabean meal or fishmeal is desirable (Gadzirayi, 2012).

Plants and their metabolites, known as bioactive compounds, play a key role because of their feed additive attributes (Uduji *et al.*, 2020). These bioactive compounds, such as carotenoids,

Copyright: Ikpe et al. Open Access. This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

flavonoids and essential oils help to maintain animal's health and productivity, and to produce safe and healthy chicken eggs (Ao *et al.*, 2011 and Ahaotu *et al.*, 2019c). The primary mode of action of these active ingredients is inhibition of pathogenic microbes (Osuagwu *et al.*, 2020) and endotoxins in the gut and enhanced pancreatic activity, resulting in better nutrient metabolism and utilization (Embuscado, 2015; Naazieand Akoto, 2011).

Moringa oleifera belongs to the family *Moringaceae* is widespread throughout the tropics. It is a small tree with sparse foliage, white flowers and long pods, often planted in farms and compounds or used as fence, especially in northern Nigeria (Ahaotu *et al.*, 2018a). It is known as Horse – radish tree, Oil of Ben tree and zogallagandi (in Hausa), ewe – igba (Yoruba) and okwe oyibo (Igbo). Its usual habitat is in farms and compounds.

In Nigeria, *Moringa* trees are distributed both in the south and north where it features as a multipurpose tree providing items of food, medicine, household, water treatment, farming system among other uses such as revenue generation through products formulation (Ahaotu *et al.*, 2018b). The tree is regarded as a miracle and life-saving resource with enormous nutritional and medicinal benefits readily providing the needs of local populace.

Reports indicated that synthetic growth enhancers and supplements in poultry nutrition are expensive, usually unavailable and may as well possess adverse effects in birds and humans (Ghazalah and Ali2008; Lee *et al.*, 2008). Probiotics for example are viable single or mixed cultures of microorganisms that when administered to animals would exert beneficially affects by improving nutrient utilization and enhance the beneficial properties of the gastro-intestinal microflora (Kyriakis *et al.*, 1999; Lee *et al.*, 2008), however, improper storage and administration, indiscriminate use of antibiotics may affect the desired effects of probiotics. It has therefore become imperative to carefully source stable, cheap, locally available and highly nutritive feed ingredients needed for poultry feed formulations.

Phytogetic feed additives (PFAs) are plant based components that are of nutritive and medical values and which seem efficient in meeting up the current challenges of livestock feeds (van der Klis, 2015). For a long time, plant

proteins are identified as cheap and are most naturally available in the tropics (Esonu *et al* 2006; Iheukwumere *et al.*, 2008; Gadziriyia *et al.*, 2012). Leaf meals obtained from many tropical plants such as *Moringa* are affordable sources of protein and micronutrients in human diets, and have therefore stimulated a lot of interest in poultry nutrition research (Abou-Elezz *et al.*, 2011; Olugbemi *et al.*, 2012).

However, most studies conducted on moringa plant seems to have been on the leaves (Du *et al.*, 2007; Kasolo *et al.*, 2010; Nuhu, 2010; Olugbemi *et al.*, 2010; Moyo *et al.*, 2011; Gadzirayi *et al.*, 2012; Mukumbo *et al.*, 2014) with little or no research on moringa pods that is often allowed to mature into seeds for propagation or sometimes thrown away together with uneatable branches.

The plant possesses many valuable properties which make it of great scientific interest. These include the high protein content of the leaves twigs and stems, the high protein and oil contents of the seeds, the large number of unique polypeptides in seeds that can bind to many moieties, the presence of growth factors in the leaves (Moreki and Gabanakgosi, 2014). The study however viewed the following objectives to investigate the effects of including *Moringa oleifera* pod meal on egg quality and performance of Shaver brown layer birds, determine the levels of inclusion of *Moringa oleifera* pod meal in diets of Shaver brown layer birds and assess the effects of essential nutrients and antioxidant-enriched *Moringa* pods on production performance.

In Nigeria, the very high cost of poultry feed has limited the expansion of the poultry industry as this has forced many poultry producers to folding. This has further drawn the already low intake of animal protein by Nigerians.

Layers premix is however expensive as a feed ingredients and this therefore increased the cost of poultry production. In order to arrest this trend, efforts are being directed towards the use of some unconventional and cheaper feed ingredients as additives with locally available, cheaper and quality feed ingredients. The use of *Moringa oleifera* pod meal to replace layers premix in layer diets is a possible way of reducing feed cost and producing cheaper eggs and meat for the populace.

Using *Moringa oleifera* pod meal as a feed additive in layer diets will go a long way in reducing the cost of production in poultry

enterprise thereby making animal protein more available to the populace and also improving the profitability of the poultry farmers. Also findings from this research would represent a significant contribution to science and knowledge.

Materials and Methods

Moringa oleifera pods were collected from the botanic garden of forestry department, Imo State Polytechnic Umuagwo and stored in polythene bags after shade drying and grinding for further analysis and addition to feed (Banjo, 2012). The pods were stripped off, washed, allowed to drain and spread in a well-ventilated room to dry for five days. Shade-dried moringa pods were milled into powder using a blender (National Mx-795N), sieved with a muslin cloth and stored for use.

The pod meal was analysed for chemical composition in the Department of Science Laboratory Technology, Imo State Polytechnic Umuagwo, according to standard procedures (Mehta *et al.*, 2003 and AOAC, 2005) (Table 1).

Table 1: Chemical Composition of *Moringa oleifera* Pod Meal

Chemical Composition	Proportion	Unit
Moisture	8.05	g/100 g
Crude Protein	18.98	g/100 g
Ether Extract	2.34	g/100 g
Ash	7.88	g/100 g
Crude Fibre	48.00	g/100 g
Minerals		
Sodium	805	mg/100 g
Potassium	2815	mg/100 g
Calcium	291	mg/100 g
Magnesium	251	mg/100 g
Phosphorus	9456	mg/100 g
Iron	53	mg/100 g
Sulphur	137	mg/100 g
Copper	3.1	mg/100 g
Oxalic acid	10	mg/100 g
Selenium	25.71	mg/100 g
Bioactive Compounds		
Quercetin	114	mg/100 g
Vitamins		
Vitamin A-B carotene (mg)	0.11	mg/100 g
Vitamin B-choline (mg)	423	mg/100 g
Vitamin B1-thiamin (mg)	0.05	mg/100 g
Vitamin B2-riboflavin (mg)	0.07	mg/100 g
Vitamin C-ascorbic acid (mg)	120	mg/100 g
B-carotene	2.76	mg/100 g
Vitamin E-tocopherol acetate (mg)	-	
Arginine (mg)	90	mg/100 g
Histidine (mg)	27.5	mg/100 g
Lysine (mg)	37.5	mg/100 g
Tryptophan (mg)	20	mg/100 g

Experimental Animals

Sixty (24 weeks old) Shaver brown layer birds were assigned to four treatments and three replicates with five birds each in a completely randomized design. Four levels substitution levels (0, 5.00, 10.0, and 02.0g MOPM/kg) were added to the four diets.

Data Collection

A daily feed allowance of 100g per bird was offered (Table 2). Feed offered, feed refused, feed intake and mortality were recorded daily and tabulated cumulatively for FCR every week. The total number of eggs laid by each replicate group of birds was determined on daily basis at 1200 hours and 1800 hours. Also daily egg production was recorded from each experimental unit separately to calculate various parameters, including egg weight, feed per dozen.

Table 2. The Performance Characteristics of Shaver Brown Pullet Birds Fed *Moringa oleifera* Pod Meal as Feed Additive.

Parameters	T ₁	T ₂	T ₃	T ₄	SEM
Mean live weight at 19 weeks (Kg)	1.615 ^a	1.555 ^b	1.575 ^b	1.570 ^b	0.04*
Mean live weight 24 weeks old (Kg)	2.115 ^a	1.975 ^b	1.970 ^b	1.960 ^b	0.06*
Mean daily weight Gain (g)	55.3 ^c	54.2 ^c	53.8 ^c	50.6 ^b	0.55*
Mean daily feed intake (g)	165.73 ^a	166.14 ^b	166.32 ^b	169.82 ^b	0.45*
Feed conversion ratio	4.43	5.01	5.34	5.46	0.08 ^{ns}
Feed cost/Kg weight gain (N)	179.24	177.84	176.44	175.04	0.10 ^{ns}
Mortality	1	1	1	4.0	0.08 ^{ns}

abc: Means with same row having different superscripts were significantly different ($P < 0.05$).

Data Analysis

Data collected was analysed through one-way ANOVA (Steel *et al.*, 1997) using PROC GLM in SAS software (SAS Inc. 9.4). Significant means was separated through Duncan's multiple range tests (Gordon and Gordon, 2004).

Results and Discussion

Results of feed consumption and weight gain of shaver brown layer birds fed with *Moringa oleifera* pod meal as feed additive, Probiotic and Vitamin E and Selenium were followed by carcass parts measurements of shaver brown pullet birds fed *moringa oleifera* pod meal as feed additive (Table 3).

Table 3. Carcass Parts Measurements of Shaver Brown Pullet Birds Fed *Moringa oleifera* Pod Meal as Feed Additive.

Parameters	T ₁	T ₂	T ₃	T ₄	SEM
Shank length (cm)	16.1 ^c	15.4 ^b	15.3 ^b	14.3 ^a	0.12*
Wing length (cm)	28.5	28.4	28.3	27.8	0.05 ^{ns}
Body length (cm)	49.4 ^c	49.3 ^c	49.2 ^c	45.4 ^a	0.03*
Body height (cm)	44.1 ^c	43.5 ^c	43.1 ^c	41.5 ^a	0.17*
Thigh length (cm)	13.9 ^b	13.7 ^b	13.3 ^b	12.9 ^a	0.07*
Leg length (cm)	11.6 ^a	10.9 ^b	10.8 ^b	10.6 ^b	0.07*
Heart girth (cm)	37.9	37.5	37.5	38.3	0.15 ^{ns}
Head Circumference(cm)	12.6 ^a	12.7 ^a	12.9 ^a	13.8 ^b	0.09*
Live weight at 24 weeks(gm)	2115 ^d	1980 ^c	1975 ^c	1860 ^a	0.09*
Dressed carcass Weight (gm)	1715 ^d	1580 ^c	1500 ^b	1480 ^a	0.08*
Eviscerated weight (gm)	1300 ^c	1280 ^a	1280 ^a	1250 ^b	0.02*

abc: Means within same row, having different superscripts are significantly different ($P < 0.05$)

The significantly ($P < 0.05$) lower live weight, fully dressed carcass weight and eviscerated weight of birds on diet T₄ could be attributed to high level of 20% MOPM in the diet. This has led to the reduction in live weight gain and carcass weight of the tested animals as reported by (Zanu *et al.*, 2012 and Ziggers, 1997). The author observed that there was a general depression in eviscerated weight by birds fed higher levels of MOPM diet. The average live weight at 24th week reported in this study are higher than those reported by Hayse and Marion (1973) who observed that mean live weights at 24 weeks pullet layer birds were below 1800gms. The lower values recorded in shank lengths, body lengths body heights and thigh lengths in treatments T₃ and T₄ was as a result of higher inclusion levels of MOPM in the diets.

It was noticed that chickens in T₁ had significantly higher ($P \leq 0.05$) body weight gain than those in the other 3 treatment groups. However, chickens in T₂ had higher ($P \leq 0.05$) body weight gain than those in treatment groups T₃ and T₄ respectively. There was no significant difference in weight gain of chickens in T₃ compared with chickens in group T₄ respectively.

The significantly ($P < 0.05$) depressed weight gain and feed conversion ratios of birds on diet T₄ was attributed to higher crude fiber in the diet (Ali

and Ali 2002; Apple *et al.* 2005). These authors further reported that high crude fibers in the diet affect growth rate. The improved performance of birds on 5%, 10% and up to 20% MOPM in this phase agreed with literature findings that MOPM improves the nutritive value of diets. Obi and Sonaiya (1995); FLD, (1981); Robert (2005); Sonaiya (1995) and Roland *et al.* (2004) stated that an essential practical consideration in evaluating a ration for animals is the cost of the feed in terms of returns obtained for the products. The 4.4% mortality rate at diet T₄ observed between 19th and 24th week (Table 4) was lower than the results of Ali and Ali (2002), Leeson and Caston (1999) who reported mortality rate of 6% for layer pullets with live weight up to 1.5kg.

Table 4. Egg Laying Performance of Shaver Brown Layer Birds Fed *Moringa oleifera* Pod Meal as Feed Additive From Point of Lay.

Parameter	T ₁	T ₂	T ₃	T ₄	SEM
Egg circumference (gm)	19.99 ^a	18.45 ^b	13.45 ^b	13.24 ^b	0.60*
Hen day production (%)	87.96 ^d	79.80 ^c	71.63 ^c	58.0 ^b	2.21*
Egg weight (gm)	62.66 ^a	60 ^b	59.4 ^b	57.4 ^b	1.84*
Mean Egg Production /Number	58 ^c	54 ^b	50 ^b	42 ^b	1.99*

abcd: Means within same row, having different superscripts are significantly different ($P < 0.05$)

The results showed that the average egg circumferences determined were 19.99, 18.45, 13.45 and 13.24 cms for birds on diet T₁, T₂, T₃ and T₄ respectively. T₁ ($P < 0.05$) had significantly the highest value and differed significantly from all the treatments. T₄ had ($P < 0.05$) significantly the lowest value while T₂ and T₃ were similar.

Egg production of the birds differed ($P < 0.05$) significantly between T₁ and T₄. However, the highest average egg numbers of 58 was recorded for birds on T₁ and the least number 42 was for T₄. Highest percentage hen - day egg production were significantly ($P < 0.05$) higher for birds on T₁, T₂, T₃ and T₄ in that order. Birds on T₄ had the least hen - day egg production. Hen - day productions were 87.96, 79.80, 71.63 and 58.00% for birds on T₁, T₂, T₃ and T₄ respectively, while mean number of eggs laid were 58, 54, 50 and 42 for the four treatments (T₁, T₂, T₃ and T₄).

Low digestibility of diet, T₄ (20% MOLM) could have resulted in its relative higher rate of

consumption over the other diets. The high dietary fiber in the diet made its component nutrients less digestible and available to the birds (Van Soest and Robertson, 1976). Therefore, the birds ate more to satisfy their nutrient requirements. The relatively higher egg production of birds on treatment T₂, T₃ and T₄ could be attributed to the effect of mineral components in *Moringa oleifera* pod meal which improved the nutritional value of those diets (Akomaegbe, 2020). The average percentage hen – day egg production in this experiment is higher than the range of 49.56 and 55% reported by Ali and Ali (2002); Chandrasiri *et al.*, (2004); Akhtar – uz – Zaman (2006); Roland, (1998) and Merat *et al.* (1992) for exotic hybrid layers. Hen – day egg production of birds on the control diet T₁ and on diets T₂, T₃ and T₄ agreed with the minimum of 65% recommended by Henken *et al.* (2002); Chineke (2001) and Oladunjoye *et al.* (2002).

The sizes of eggs obtained in this study were above the standard egg weights of 58 gm reported in the literature (Ali 2002; Austic and Nesheim 2003; Demeke 2006 and Doran *et al.* 1990) for exotic breeds of layers. The relatively low values of eggs obtained from birds on diet T₄ is in agreement with Austic (2005); Banerjee (2002); Merat *et al.* (1992); Parsons *et al.* (1998) and Roland (1998) who reported that higher crude fibre rations decrease egg production just like water deprivation. The superior egg circumference qualities obtained from treatment diets T₂ and T₃ support the findings of Reid *et al.* (2004) and Singhet *et al.*, (2002) that MOPM positively influences nutrient utilization, thereby impacting on egg production.

Herb extracts have been reported to significantly improve body weight gain, feed conversion ratio as well as layer carcass dressing percentages (Omar *et al.*, 2016). Photogenic feed additives containing essential oils, spices and saponins have been reported to significantly influence nutrient digestibility in layer birds. Plant derived feeds are believed to possess antioxidant properties that improve nutrient absorption by cells, strengthen cellular defense and minimize damages by oxidative stress which consequently leads to improvement in health status of animals (van der Klis, 2015).

Results showed that *Moringa oleifera* pod meal inclusion in layer diets had decreased feed intake but improve live weight gains in layer chickens.

The least feed intake as well as the highest weight gain was noticed in *moringa* pod treated group as compared to other treatment groups and the control thus suggesting *moringa* pods to be of economic value especially in terms of its probable feed conversion efficiency. Lannaon (2007) reported high improvement in daily weight gain, final weight gain and profit on the performance of Shaver brown layers given *Moringa oleifera* pod meal.

Worthy of noting also was work of Du *et al.*, (2007) who evaluated the effect of dietary supplementation of *Moringa oleifera* on growth performance, blood characteristics and immune response of Abore acre strain of broilers. It was found that increasing supplementation of *Moringa oleifera* decreases the contents of uric acids, triglycerides and albumin ratio in the serum of broilers. Hence, immune response of broilers increased significantly. Also Ahaotu *et al.*, (2019 a) and Yang *et al.*, (2007) evaluated the effect of *Moringa oleifera* on growth performance, immune function and ileum microflora in broilers.

However, *Moringa* seed pod has also been reported to be a source of activated carbon used to treat poisons in addition to its rich source of vitamins A, B and C, protein, sulphur containing amino acids, calcium, phosphorus and iron (Ahaotu *et al.*, 2019 b and Mohammed, 2013) which most likely could account for its nutritive value that led to higher weight gain observed in this study. It is also noted that the improvement in body weight gain in *moringa* treated group could be due to improved feed conversion or reduced amount of feed required to produce one unit of meat.

Probiotics on the other hand are live nonpathogenic and non-toxic microorganisms, which when administered through the digestive route favour the host's health (Guillot, 1998). The addition of probiotics especially Bactofort® and Anthox® to livestock feed were reported to improve the nutritive quality of feed and performance of animals by (Martin *et al.*, 1989).

Non-antibiotic growth promoters such as organic acids and probiotics are increasingly being produced and used for animal nutrition worldwide (Windisch *et al.*, 2008). The cost may hinder many backyard poultry farmers in especially developing nations and therefore *moringa* pods may be an excellent alternative.

One of the major reasons for increased interest in the use of probiotics in livestock in recent years

is because they are natural alternatives to antibiotics and for their growth promotion effects in especially poultry (Bajpai *et al.*, 2005). Rajmane and Ranade (1994) found that the inclusion of vitamin E and C at 150 mg/kg and 200 mg/kg in poultry diet respectively improved growth rate as well as immune response of vaccinated chickens. Furthermore, Bharali *et al.* (2003) reported that inclusion of selenium in poultry diet markedly reduced feed conversion ratio due to the fact that it significantly lowered feed intake but found it to have improved eviscerated carcass weight.

Contrary, Bhattacharya *et al.* (1982) reported that selenium yeast improved body weight and feed conversion ratio in layers. In this study however, it was found out that layers treated with vitamin E and Selenium had a high feed consumption with a concomitant least body weight gain agreeing closely to the findings of Atuahene *et al.* (2008). Vitamin E and selenium may therefore be of little or no value in enhancing body weight gain in layers.

The decrease in the feed intake was due to high density feed on account of essential amino acids, vitamins and minerals present in MOPM which meet the body requirement even with smaller intake (Abbas and Ahmed, 2012). During the experimental period best FCR and BWG was recorded in the T₃ with 10% MOPM. This might be due to rich availability of essential nutrients like amino acids, vitamins and antioxidant compounds present in *Moringa oleifera* pods (Abou-Elezz *et al.*, 2011) which affect the overall health, production and FCR in experimental layers. The increase in relative giblet weights can be attributed to the bioactive compounds (carotenoids, flavonoids) of *Moringa* pods meal, which interact with the metabolism and enhance the productive performance by improving digestibility. Some other studies also reported that *Moringa oleifera* supplementation affect the giblet relative weight (Glade and Sist, 1998). Similarly it has been reported in other studies that *Moringa* supplementation showed positive impact on FCR of broiler birds (Ahaotu *et al.*, 2019).

Conclusion

It was concluded that MOPM at 10% inclusion in layers feed improved egg production and body weight. It is recommended that more extensive work with larger number of birds from point of lay to the end of lay and higher levels of MOPM be carried out for more authentic conclusion and

recommendation. The pod meal also had relative economic efficiency and could serve as an excellent alternative to commercially available growth promoters.

Reference

- Abbas TE (2013). The use of *Moringa oleifera* in poultry diets. Turk. J. Vet. Anim. Sci. 37: 492-496.
- Abbas TE and Ahmed (2012). The use of *Moringa oleifera* in poultry diets. Turkish Journal of Veterinary and Animal Science. 11-40.
- Abou-Elezz FMK, Sarmiento-Franco L, Santos-Ricalde R and Solorio-Sanchez JF (2011). Nutritional effects of dietary inclusion of *Leucaena leucocephala* and *Moringa oleifera* leaf on Rhode Island Red hens, performance. Cuban J. Agric. Sci. 45: 163-169.
- Ahaotu EO, Aturuchi W, Unagu A and Uzegbu HO (2019a). Replacement value of African Bread Fruit Seed Meal (*Treculia africana*) for Maize on the Performance of Starter Broilers. Sustainability, Agri, Food and Environmental Research, 7(2): 32-43.
- Ahaotu EO, Abiola MO and Madu WC (2019b). Anti-microbial Efficacy of *Moringa oleifera* Leaves Extract Against *Escherichia coli* and *Staphylococcus aureus*. Journal of Microbiology, Immunology and Biotechnology. 6: 01-04
- Ahaotu EO, Amajioyi N and Okorie KC (2019c). Sustainability of Guinea Fowl (*Numidia meleagris*) layers fed varying levels of *Centrosema pubescent* leaf meal. Sustainability, Agri, Food and Environmental Research. 7 (1): 1-10
- Ahaotu EO, Uwalaka RE, Edih MC and Ihiaha PO (2018a). Purification of Water using *Moringa oleifera* Horse radish) Leaf Powder and Seeds Flour. Annals of Agric. Sci., Moshtohor, Egypt. 56(4): 1027 – 1030
- Ahaotu EO, Uwalaka RE, Edih MC and Ihiaha PO (2018b). Extraction of Oil, Biogas and Biodiesel from *Moringa oleifera* Seeds. Annals of Agric. Sci., Moshtohor, Egypt. 56(4): 1021-1026.
- Ahaotu EO, Nwoye EO and Lawal Muhammad (2019). Biochemical Constituents, Economics of Production and Performance of Starter Broilers Fed Diets Supplemented with Natural Feed Additives. Int. Journal of Animal and Veterinary Sciences. 06 : 01-05
- Akhtar-Uz-Zaman M (2006). Egg Production

- Performance of different breed/breed Combinations of chicken in semi-scavenging system under PLDP. Ph.D Dissertation. The Royal Vet. and Agric. Uni, Denmark.
- Akomaegbe C (2020). Response of Inclusion of Dried Horse Radish (*Moringa oleifera*) Pod Meal as Feed Additive on Egg Laying and Performance of Shaver Brown Birds. Student Project. Department of Animal Production and Health Technology, Imo State Polytechnic Omuma Campus, Nigeria. 67 pp.
- Ali MS (2020). Study on the effect of energy diets to laying hens under the rural condition of Bangladesh. The Royal Vet. and Agric. Uni. Press; Denmark. 498 pp.
- Ali MS and Ali KO (2002). Study on the effect of hydrolysed feather meal supplementation to laying hen under the rural condition of Bangladesh. J. Roy. Vet. Agric. Uni. Denmark. 86 pp.
- Aniebo CM (2013). Effects of *Moringa oleifera* leaf meal on the performance and blood haematological and chemistry of starter broilers. Int. Journal of Food Agriculture and Veterinary Science ICD: 38- 44.
- Ao X, Yoo JS, Zhou TX, Wang JP, Meng QW, Yan L, Cho JH and Kim IH (2011). Effects of fermented garlic powder supplementation on growth performance, blood profiles and breast meat quality in broilers. Livest.. Sci. 141: 85-89.
- A.O.A.C. (2005). Association of Official Analytical Chemists. Official Methods of Analysis. Washington D.C. USA. 69-88 pp
- Apple JK, Boger CB, Brown DC and Johnson ZB (2005). Effect of feather meal on live Animal performance, carcass quality and composition of growing finishing Swine. J.of Anisci.;Univ of Arkansas, Fayette Ville, U.S.A
- Atuahene CC, Attoh-Kotoku V, Fosu KD, Amissah SE, Sarfo FK and Mensah JJ (2008). Preliminary study of the effect of feeding *Moringa Oleifera* leaf meal as a feed ingredient on the growth performance of broiler chickens. Proceedings of the Ghana Animal Science Association held at the University of Education, Winneba, Mmpong Campus, 28th and 29th Aug., 2008. pp.72-75.
- Austic RE (2005). Feeding Poultry in Hot Climates. Revised Edition, CRC Press, U.S.A 698 pp
- Austic RE and Nesheim MC (2002). Poultry Production. Revised Edition, Lea and Febiger Press, U.S.A 793 pp.
- Bajpai M, Pande A, Tewari SK and Prakash D (2005). Phenolic contents and antioxidant activity of some food and medicinal plants. Int. J. Food Sci. Nutr., 56: 287-291.
- Banerjee GC (2002). Poultry; Revised Edition. Oxford and IBH Publishing Coy. PVT LTD, New Delhi pp 168-172.
- Banjo OS (2012). Growth and performance as affected by inclusion of *Moringa oleifera* leaf meal in broiler chicks diet. J. Biol. Agri. Healthcare. 2: 2224-3208.
- Bharali R, Tabassum J and Azad M (2003). Chemo-modulatory Effect of *Moringa oleifera*, Lam, on hepatic carcinogen metabolising enzymes, antioxidant parameters and skin papillo-magenesis in mice. Asia Pec. J. Cancer Prev., 4: 131-139.
- Bhattacharya SB, Das AK and Banerji N (1982). Chemical investigations on the gum exudate from sajna (*Moringa oleifera*). Carbohydr. Res., 102: 253-262.
- Chandrasiri AD, Gunaratne S, Wickramaratne SN and Roberts JA (2004). The egg and meat production potential of laying hens. In: sustainable Animal Production. Proc. Of Ani. Sci. Congr. Bali; Indonesia. 73-75.
- Chineke CA (2001). Interrelationships existing between body weight and egg production traits in Olympia black layers. Nig. J. Anim. Prod., 28: 1-8.
- Demeke S (2006). Study on egg production of white leghorn under intensive, Semi-intensive and rural household conditions in Ethiopia. Livestock Research for Rural Development. 8 (2): 1-55.
- Doran BH, Quesenberry JH, Krueger WF and Beadley JW (1990). Response of thirty Egg type stock to four layer diets differing in protein and caloric levels. Poult. Sci. 68: 108-1089.
- Du PL, Lin PH, Yang RY and Hsu JC (2007). Effect of dietary supplementation of *Moringa oleifera* on growth performance, blood characteristics and immune response in broilers. Journal of Chinese Society of Animal Science, 36(3): 135-146.
- Embuscado ME (2015). Spices and herbs: Natural

- sources of antioxidants - a mini review. J. Func. Foods. 1-9 pp
- Esonu BO, Opara MN, Okoli IC, Obikaonu HO, Udedibie C and Iheshiulor OOM (2006). Physiological response of laying birds to Neem (*Azadirachta indica*) leaf meal-based diets; body weight, organ characteristics and hematology. Online J. Health. App. Sci 2, 4 http://www.ojhas.org/issue_18/2006-2-4.htm).
- FEDERAL LIVESTOCK DEPARTMENT {FLD} (1981). The cost of production and guaranteed minimum prices for selected livestock production in Nigeria. Centre for Agriculture and Rural Development (CARD), Department of Agricultural Economics, University of Ibadan, Ibadan. 173 pp.
- Gadzirayi CT, Masamha B, Mupangwa JF and Washaya S (2012). Performance of Broiler chickens fed on mature *Moringa oleifera* leaf meal protein supplement soyabean meal. Int. J. Poult. Sci. 11(1): 5-10.
- Ghazalah AA and Ali AM (2008). Rosemary leaves as dietary supplement for growth in broilers. International Journal of Poultry Science, 7(3): 234-239.
- Glade MJ and Sist MO (1998). Dietary yeast culture supplementation enhances urea recycling in equine large intestine. Nutritional Reproduction International, 37: 11-17.
- Gordon SP and Gordon FS (2004). Contemporary Statistics: A Computer Approach. McGraw - Hill Publishers, U.S.A. 98-112pp.
- Grashorn MA (2010). Use of phytobiotics in broiler nutrition: an alternative to in feed antibiotics. J. Anim. Feed. Sci. 19: 338-347.
- Guillot JF (1998). Les probiotiques en alimentation animale. Cahiers Agricultures, 7: 49-54.
- Hayse PL and Marion WW (1973). Eviscerated yield, component parts and meat skin and bone ratios in the pullet chicken. Poult. Sci., 52: 718-722.
- Henken AM, Croote AM and Van-derhel W (2002). The effect of environmental temperature on immune response, metabolism and laying performance of pullets. Poult. Sci., 82(4): 59-67.
- Iheukwumere FC, Ndubuisi EC, Mazi EA and Onyekwere MU (2008). Performance, nutrient utilization and organ characteristics of broilers fed cassava leaf meal (*Manihot esculenta* Crantz). Pak. J. Nutr. 7: 13-16.
- Kakengi AMV, Kaijage JT, Sarwatt SV, Mutayoba SK, Shem MN and Fujihara T (2007). Effect of *Moringa oleifera* leaf meal as a substitute for sunflower seed meal on performance of laying hens in Tanzania. Livestock Research for Rural Development, 19(8): 120.
- Kasolo JN, Bimenya GS, Ojok L, Ochieng J and Ogwal-Okeng JW (2010). Phytochemicals and uses of *Moringa oleifera* leaves in Ugandan rural communities. J. Med. Plant Res., 4: 753-757.
- Kyriakis SC, Tsiloyiannis VK, Vlemmas J, Sarris K, Tsinas AC, Alexopoulos C and Jansegers L (1999). The effect of probiotic LSP, 122 on the control of post weaning diarrhoea syndrome of piglet. Research in Veterinary Science, 67: 223-228.
- Lannaon WJ (2007). Herbal Plants as Source of Antibiotics for Broilers. Agriculture Magazine. 11(2): 55.
- Lee NK, Yun CW, SW, Chang HI, Kang CW and Paik HD (2008). Screening of *Lactobacilli* derived from chicken faeces and partial characterization of *Lactobacillus acidophilus* A12 as an animal probiotics. Journal of Microbiology and Biotechnology, 18: 338-342.
- Leeson S and Caston LJ (1999). Response of laying hens to diets varying in crude protein and available phosphorus. J. Appl. Poult. Res. 5: 289-296.
- Martin SA, Nisbet BJ and Dean RG (1989). Influence of a commercial yeast supplement on the in vitro ruminal fermentation. Nutritional Reproduction International, 40: 395-403.
- Mehta K, Balaraman R, Amin AH, Bafna PA and Gulati OD (2003). Effect of fruits of *Moringa oleifera* on the lipid profile of normal and hyper-cholesterolaemic rabbits. J Ethnopharmacol, 86 (2): 191-195.
- Merat PG, Coquerell G and Durand L (1992). Phenotypic correlations of egg characters with growth and laying performance of hens in two lines producing brown eggs and their daughters. Archiv fur Geflugelkunde, 56:99-105.
- Mohammed IJ (2013). Medicinal and dietary roles of *M. stenopetala* in South Ethiopia. African Journal of Agricultural Science and

- Technology, 1(1): 1-6.
- Moreki JC and Gabanakgosi K (2014). Potential use of *Moringa oleifera* in poultry diets. *Global J. Anim. Sci. Res.* 2(2): 109-115.
- Moyo B, Masika PJ, Hugo A and Muchenje V (2011). Nutritional characterization of *Moringa (Moringa oleifera Lam.)* leaves. *African Journal of Biotechnology*, 10(60): 12925-12933.
- Mukumbo FE, Maphosa V, Hugo A, Nkukwana TT, Mabusela TP and Muchenje V (2014). Effect of *Moringa oleifera* leaf meal on finisher pig growth performance, meat quality, shelf life and fatty acid composition of pork. *South African Journal of Animal Science.* 44(4): 389-400.
- Naazie A and Akoto KD (2011). The feeding value of moringa (*Moringa oleifera*) leaf meal in broiler diets. *Proceedings of the 17th Biennial Conference of the Ghana Society of Animal Production (GSAP)*, pp. 104-107.
- Nkukwana TT, Muchenje V, Pieterse E, Masika PJ, Mabusela TP, Hoffman LC and Dzamab K (2014). Effect of *Moringa oleifera* leaf meal on growth performance, apparent digestibility, digestive organ size and carcass yield in broiler chickens. *Livest. Sci.*, 161: 139-146.
- Nuhu F (2010). In: Effect of *Moringa oleifera* leaf (MOLM) on Nutrient Digestibility, Growth, Carcass and Blood Indices of Weaner Rabbits Kwame Nkrumah University of Science and Technology Kumasi, Ghana (Ph.D. thesis).
- Obi O and Sonaiya EB (1995). Gross margin analysis of small holder rural Poultry Production in Osun State. *Nig. J. Anim. Prod.* 22: 95-107.
- Odoemelum S, Simeon-Ahaotu VC, Patricio De Los Ríos and Ahaotu EO (2020). Effects of Frangipanni (*Plumeria Rubra*) Leaf Meal as Feed Additive on the Performance and Egg Laying Index of Hy-Line Brown Birds. *International Journal of Research in Agriculture and Forestry.* 7(2): 1-6
- Oladunjoye IO, Amao OA and Adeleke JA (2002). Effects of bread fruit Meal on laying performance, Egg qualities and Serum chemistry of chickens. *Trop. Anim. Prod. Invest.* 5:129-136.
- Olugbemi TS, Mutayoba SK and Lekule FP (2012). Effect of *Moringa oleifera* inclusion in cassava based diet fed to broiler chickens. *International Journal of Poultry Science*, 9: 363-367.
- Olugbemi TS (2010). *Moringa oleifera* leaf meal as a hypocholesterolemic agent in laying hen diets. *Livestock Research for Rural Development*, 22(4): 1-12
- Omar AJ, Hejazi A and Badran R (2016). Performance of broilers supplemented with natural herb extract. *Open Journal of Animal Science*, 6: 68-74.
- Onu PN (2011). Impact of heat treated sheep manure-based diets with or without exogenous enzyme on nutrient digestibility and economics of production of finisher broilers. *International Journal of Science and Nature.* 2(1): 1-5
- Osuagwu CO, Anosike C, Ahaotu EO and Oko EC (2020). Factors Influencing Farmer Adoption of Improved Poultry Production Practices in Ahiazu Mbaise Local Government Area of Imo State, Nigeria. *Annals of Microbiology and Infectious Diseases.* 3(3): 4 - 11.
- Parsons CM, Koelkebeck KW, Zhang Y, Wang X and Leeper RW (1998). Effect of dietary protein and added fat levels on performance of young laying Hens. *J. Appl. Poult. Res.* 2:214-220.
- Rajmane BV and Ranade AS (1994). Medicinal and dietary roles of *M. stenopetala* in South Ethiopia. *African Journal of Agricultural Science and Technology*, 1(1): 1-6.
- Reid WM, Pesti AM, Hammarlund MA and Vohra P (2004). Raising health Poultry under primitive conditions. CVM Press, Seattle, U. S. A 103 pp.
- Roberts JA (2005). Utilization of feed resources by small holders in the villages of developing Countries. *Proc. Workshop. The Danish Agric. And Rural Develpt.* 311-335.
- Roland DA (1998). Egg shell quality: Effect of dietary manipulations of protein, amino acids, energy and calcium in aged hen on egg weight, shell weight, shell quality and egg production. *Poult. Sci.* 69:2038-2046.
- Roland DA, Rao SK, Bryant MM and Self J (2004). Econometric Nutrition: Maximizing profit in commercial leghorns by optimizing feeding method, total sulphur amino acid intake and environmental temperature. *Degussa Tech. Symp.* 47-57.
- Singh RP, Murthy KN and Jayaprakasha GK (2002).

- Studies on the antioxidant activity of pomegranate (*Punica granatum*) peel and seed extracts using *in vitro* models. *Journal of Agricultural and Food Chemistry*, 50(1): 81-86.
- Sonaiya EB (1995). Feed Resources for small holder poultry in Nigeria. *World Anim. Rev.* 1: 25-33.
- Steel RGD, Torrie JH and Dickey DA (1997). Principles and Procedures of Statistics. A Biometrical Approach. 3rd ed., McGraw Hill Book Co., New York, USA. Pp154-168.
- Uduji MA, Uche MI, Simeon-Ahaotu VC, Patricio De Los Ríos and Ahaotu EO (2020). Effects of Frangipanni (*Plumeria Rubra*) Flower Meal as Feed Additive on the Performance and Egg Laying Index of Isa Brown Birds. *International Journal of Res. Agri. and Forestry*. 7(3): 1-7.
- Van der Klis DJ (2015). Counteracting heat stress in poultry production. *Poultry International*, pp. 273-275.
- Van soest PH and Robertson JB (1976). Chemical and Physical properties of dietary fiber. *Proc. Miles. Symposium*. 176, June. 14, 1976. Delhouse University Halifax NovaCotia.
- Windisch W, Schedle K, Plitzner C and Kroismayr A (2008). Use of phytogenic products as feed additives for swine and poultry. *Journal of Animal Science*, 86 (E. Suppl.): E140-E148.
- Yang RY, Chang LC, Hsu JC, Weng BBC, Palada MC, Chadha ML and Levasseur V (2007). Nutritional and Functional Properties of *Moringa oleifera* Leaves- From Germplasm, to Plant, to Health. in: *Moringa Leaves: Strategies, Standard and Market for a Planning*, ABU, Zaria, Nigeria, Pp. 1-5.
- Zanu HK, Asiedu P, Tampuori M, Asada M and Asante I (2012). Possibilities of using *Moringa oleifera* leaf meal as a partial substitute for fishmeal in broiler chickens diet. *Online J. Anim. Feed Resources*, 2(1): 70-75.
- Ziggers D (1997). *The poultry production guide*. Misset Press, Netherlands. 1200 pp.
